

RAPAMYCIN PEPTIDES CONJUGATES: SYNTHESIS AND  
USES THEREOF

**TECHNICAL FIELD**

This application relates to cell cycle inhibitors.  
5 More particularly the invention relates to the synthesis of rapamycin peptides conjugates and their use in treating disorders related to cell division.

**BACKGROUND OF THE INVENTION**

Cancer drug discovery is one of the most rapidly  
10 changing areas of pharmaceutical research. Most anticancer agents that are approved for clinical use are molecules which damage deoxyribonucleic acid (DNA), block DNA synthesis indirectly through inhibition of nucleic acid precursor biosynthesis or disrupt hormonal  
15 stimulation of cell growth (Sielecki, T.M. et al. *J. Med. Chem.* 2000, 43(1), 1-18). There has been a recent shift of emphasis towards novel mechanistic targets that has emerged as a direct consequence of the intense study of the underlying genetic changes associated with the  
20 cancerous state. The high frequency of mutations in cancer cells which results in altered cell cycle regulation, in conjunction with aberrant expression of cyclin dependent kinases (CDKs) and growth signal transduction, conferring a proliferative advantage,  
25 indicates that many of these aberrant mechanisms may be strategic targets for cancer therapy. An increasing body of evidence has shown a link between tumor development and CDK related malfunctions. Over expression of the cyclic regulatory protein and subsequent kinase

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hyperactivity have been linked to several types of cancers. The process of cell division has been amply studied but the molecular mechanisms that regulate the cell cycle have only been elucidate in the last two  
5 decades. The phases of the cell cycle are: The rest phase,  $G_0$ , active protein synthesis in preparation of cell division occurs in the  $G_1$  phase. During the  $G_1$  phase the volume of the cell increases. After the  $G_1$  phase the cells enter the S phase in which the DNA is replicated. The S  
10 phase is followed by another gap phase,  $G_2$ , during which DNA replication is completed. The last phase is the mitosis or M phase in which the cells divide (Muhtasib, H.G. et al. *Curr. Cancer Drug Targets* 2002, 2, 309-336).

Rapamycin (Sirolimus, Rapamune, 1,18-dihydroxy-12-  
15 [2(4-hydroxy-3-methoxy-cyclohexyl)-1-methyl-ethyl]-19,30-4-aza-tricyclo[30.3.1% 4,9 &] hexatriaconta-16,24,26,28-tetraene-2,3,10,14,20-pentaone) with a molecular formula of  $C_{51}H_{79}NO_{13}$  and molecular mass of 913.6 Da was isolated in 1975 from the bacteria strain *Streptomyces hygroscopicus*  
20 found in a soil sample on Ester Island (Sehgal, S.N. et al. *J. Antibiot.* 1975, 28, 721 and Sehgal, S.N. et al. *J. Antibiot.* 1975, 28, 727). Rapamycin has potent antimicrobial, immunosuppressant and antitumor properties. It inhibits the translation of key mRNAs of  
25 proteins required for the cell cycle progression from  $G_1$  to S phase by binding intracellularly to the immunophilin FK506 binding protein FKBP12 and the resultant complex inhibits the protein kinase activity of a protein kinase termed mammalian target of rapamycin (mTOR). The  
30 inhibition of mTOR, in turn blocks signals to two separate downstream pathways which control the

translation of specific mRNA (40S ribosomal protein S6 kinase P70<sup>S6K</sup>) required for cell cycle traverse from G<sub>1</sub> to S phase (Wiederrecht, G.J. et al. *Prog. Cell.Cycle. Res.* 1995, 1, 53-71).

5       The poor aqueous solubility and chemical stability of rapamycin precluded its clinical development as an anticancer agent. Recently a series of rapamycin analogs with improved aqueous solubility and stability have been synthesized and evaluated. CCI-779 (Wyeth Ayerst, PA,  
10   USA), a soluble ester analog of rapamycin is selected for development as an anti cancer agent based on its prominent antitumor profile and favourable pharmaceutical and toxicological characteristics in preclinical studies (Huang, S. et al. *Curr. Opin. Investig. Drugs* 2002, 3,  
15   295-304). CCI-779 has demonstrated significant inhibitory effects both in vivo and in vitro (various cell lines lines with IC<sub>50</sub> values of < 10<sup>-8</sup> M). Its cytostatic properties results from the inhibition of translation of several key proteins that regulate the G<sub>1</sub> phase of the  
20   cell cycle. Similar to rapamycin, CCI-779 is hypothesized to form a complex with the intracellular cytoplasmic protein FK506 binding protein -12 (FKBP) that binds to mTOR resulting in the inhibition of key signaling pathways involved in the G<sub>1</sub> phase of the cell cycle and  
25   thereby checks the progression from G<sub>1</sub> to S phase. Studies have shown that CCI-779 is able to penetrate the blood brain barrier as it has aqueous solubility and is highly lipophilic. Phase I and II studies have shown that CCI-779 is associated predominantly with skin toxicities  
30   (rash, folliculitis, prurtis, ulceration and nail changes), stomatic and asthenia (Elit, L. *Curr Opin.*

*Investig. Drugs* 2002, 3, 1249-1253 and Punt, C.J.A. et al. *Annals of Oncology* 2003, 14, 931-937).

The CDK complex activity is regulated by mechanisms such as stimulatory or inhibitory phosphorylations as well as the synthesis and degradation of the kinase and cyclin subunits themselves. Recently a link has been established between the regulation of the activity of the cyclin dependent kinases and cancer by the discovery of a group of CDK inhibitors including p27<sup>Kip1</sup>, p21<sup>Waf1/Cip1</sup> and p16<sup>Ink4/MTS1</sup>. The inhibitory activity of p27<sup>Kip1</sup> is induced by the negative growth factor TGF- $\beta$  and by contact inhibition (Nurse et al. *Nature* 1994, 372(8), 570-573). The interleukin-2 (IL-2) allows CDK activation by causing the elimination of the CDK inhibitor protein p27<sup>Kip1</sup>, which effect is prevented by rapamycin. By contrast, the CDK inhibitor p21 is induced by IL-2 and this induction is blocked by rapamycin. The activity of p21<sup>Waf1/Cip1</sup> is regulated transcriptionally by DNA damage through the induction of p53, senescence and quiescence. The tumor suppressor protein p21<sup>Waf1</sup> plays a central role in regulating eukaryotic cell-cycle progression. Through its association with G<sub>1</sub> and S phase CDK complexes it regulates activation of the retinoblastoma protein (pRb) and E2F transcription factors. Thus, selective blockade of the cyclin recruitment site would prevent recognition and subsequent phosphorylation of CDK substrates, and therefore offers a therapeutic approach towards restoration of p21<sup>Waf1</sup> like tumor suppression. Recently the octapeptide, HSKRRLIF, located C-terminal in p21<sup>Waf1</sup> which has been shown to display potent cyclic inhibitory activity due to its capacity to bind to the cyclic

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recruitment site. These proteins p27<sup>Kip1</sup>, p21<sup>Waf1/Cip1</sup> and p16<sup>Ink4/MTS1</sup>, when bound to CDK complexes, inhibit their kinase activity, thereby inhibiting progression through the cell cycle (Chen, Y.P. et al. *Proc. Natl. Acad. Sci. USA* 1999, 96, 4325-29; Zheleva, D.I. et al. *J. Peptide Res.* 2002, 60, 257-270; Atkinson, G.E. et al. *Bioorg. Med. Chem. Lett.* 2002, 12, 2501-2505; McInnes, C. et al. *Curr. Med. Chem.-Anticancer Agents* 2003, 3, 57-69.

There is therefore a need for compounds that can target the function of cell cycle suppressors such as p27<sup>Kip1</sup> and p21<sup>Waf1/Cip1</sup>.

#### SUMMARY OF THE INVENTION

The present invention relates to new rapamycin derivatives for the inhibition of cell proliferation. The compounds advantageously combine two molecular functionalities that can target the functions of two or more proteins in dividing cells and interfere with cell cycle.

In one embodiment of the invention there is provided derivatives of rapamycin in which the 42 position of rapamycin is linked to an amino acid, or an amino alcohol, or a peptide through a carbamate ester linkage. These rapamycin derivatives can be synthesized by reacting 42-O-(4-Nitrophenoxy carbonyl)rapamycin and an amino acid, or amino alcohol, or an amino peptide under basic conditions.

In a further embodiment the rapamycin derivatives can be used to inhibit the cell cycle and are therefore useful for treating cell proliferation disorders selected

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from cancer, hyperplasia, psoriasis and hyperproliferative vascular disease.

The compounds or compositions of the present invention can be released from a carrier, and the carrier  
5 can be implanted at a desired location within a patient using for example a catheter.

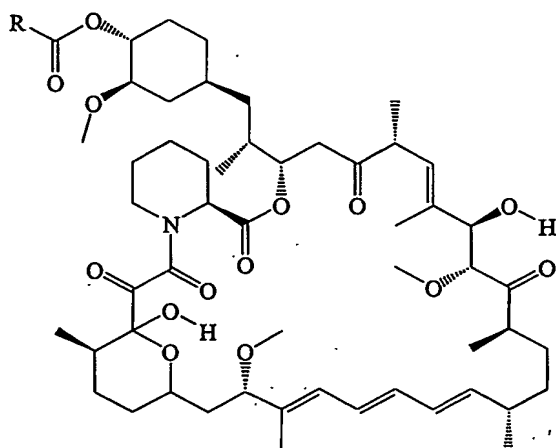
In a further embodiment there is provided a stent coated with the compound of the present invention for the treatment of a hyperproliferative vascular disease.

10 In yet a further embodiment pharmaceutical compositions comprising a compound of the invention or a pharmaceutically acceptable salt thereof, and a pharmaceutically acceptable carrier can be used as an immunosuppressant for treating an immunological condition  
15 such as autoimmune disease and host-graft disease.

Further features and advantages of the present invention will become apparent from the following detailed description.

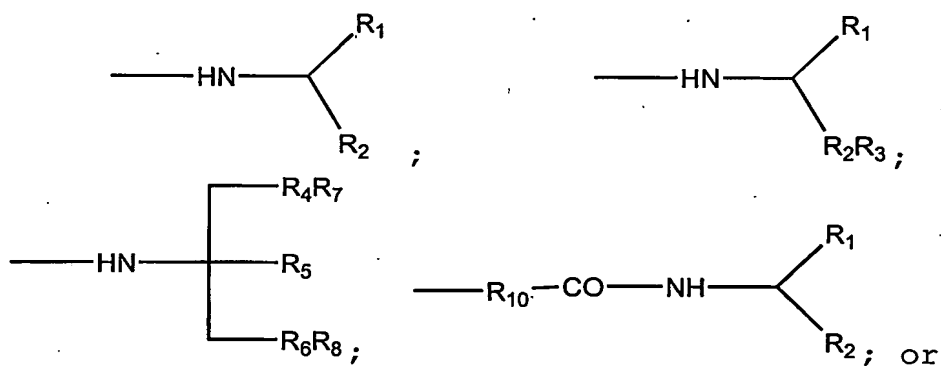
20 **DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENT**

This invention relates to the synthesis of novel rapamycin derivatives compounds useful for the inhibition of cell division for the treatment of diseases in which the inhibition of cell proliferation is desirable having  
25 the structure



wherein,

5 R is  $\text{NH}-(\text{A})_n-\text{CH}_2\text{OH}$ ;



10  $-\text{NH}-\text{R}_{11}$

A is *D* or *L* amino acid, and  $n=1-10$ , A is preferably combinations of amino acids as shown in compounds 7a-7i,  $\text{R}_1$  and  $\text{R}_2$  are each independently, hydrogen, alkyl of 1-6  
 15 carbons atoms, hydroxyalkyl of 1-6 carbon atoms, or  $\text{CO}_2\text{R}_9$ ,  $\text{R}_3$  is Ar, wherein Ar is aromatic or hetroaromatic,  $\text{R}_4$ ,  $\text{R}_5$  and  $\text{R}_6$  are each independently alkyl of 1-6 carbon atoms or hydroxyalkyl of 1-6 carbon atoms,

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R<sub>7</sub> and R<sub>8</sub> are each independently hydrogen, cycloalkyl of 1-6 carbon atoms or hydroxycycloalkyl of 3-16 carbon atoms, and

R<sub>9</sub> is alkyl of 1-6 carbon atoms,

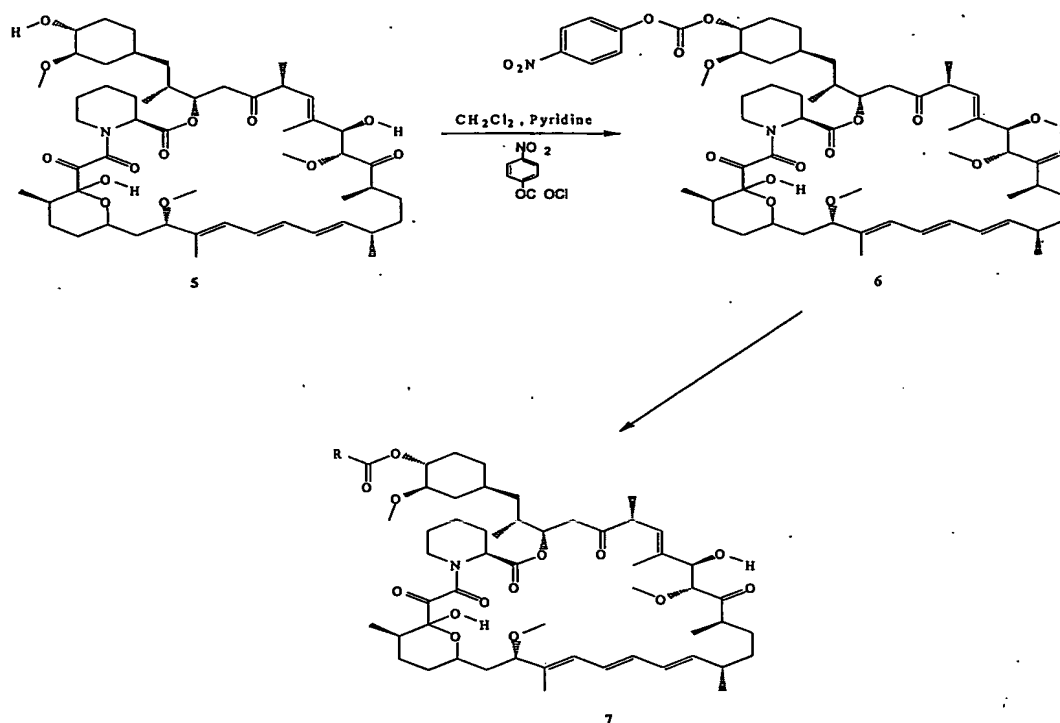
5 R<sub>10</sub> is alkyl of 1-10 carbon atoms and

R<sub>11</sub> is cycloalkoxyalkyl of 3-10 carbon atoms.

In one embodiment, amino acids and/or small peptides  
10 derivatives of the octapeptide HSKRRLIF are conjugated  
with rapamycin (formula 5). The regioselective synthesis  
of derivatives of rapamycin 5 at the 42 position, is  
achieved by conjugating the amino end of the amino acids  
and/or active peptides with 42-O-(4-Nitrophenoxy carbonyl)  
15 rapamycin (6). Compounds of general formula 7 (**Scheme 1**)  
are thereby obtained.



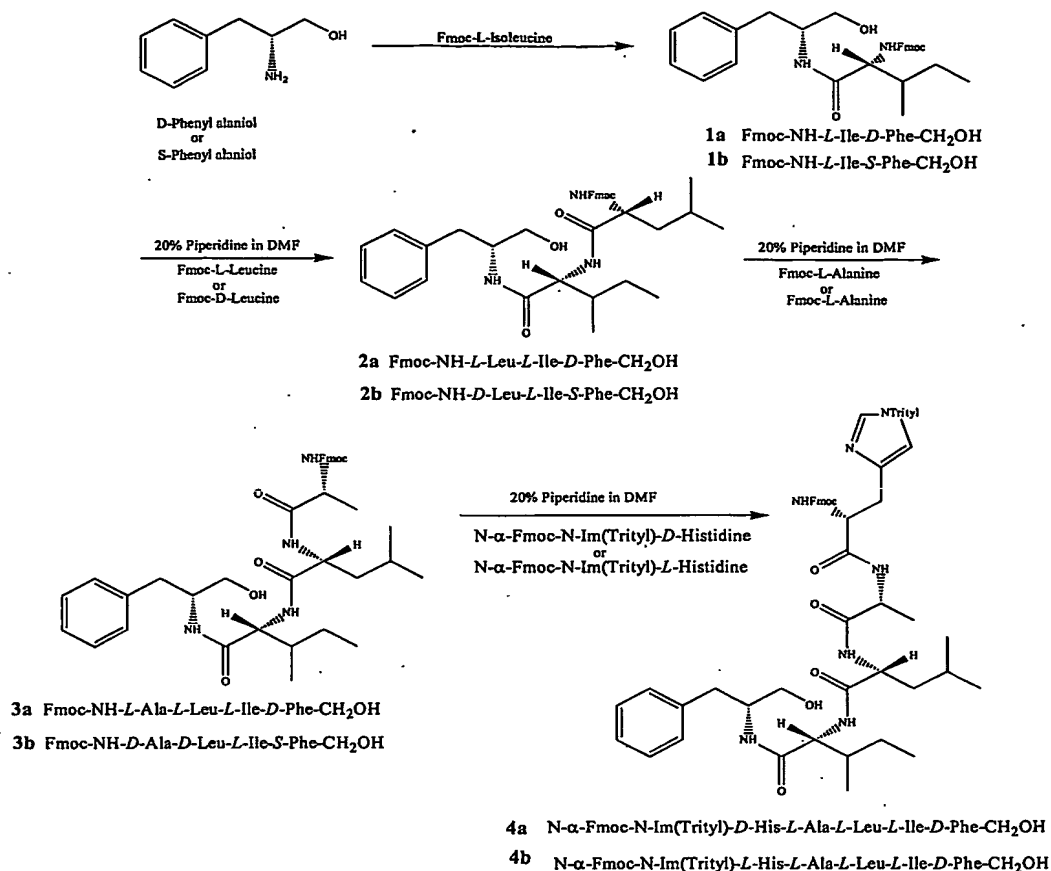
Scheme 1



The peptides conjugated to rapamycin preferably comprise amino acids from the C-terminal of the octapeptide HSKRRLIF. The amino acids at the N-terminal may differ from that of the octapeptide. Single amino acids may also be used. Examples of compounds obtained by the combination of 42-O-(4-Nitrophenoxycarbonyl) rapamycin and amino acids and/or peptides are given below (compounds 7a to 7v).

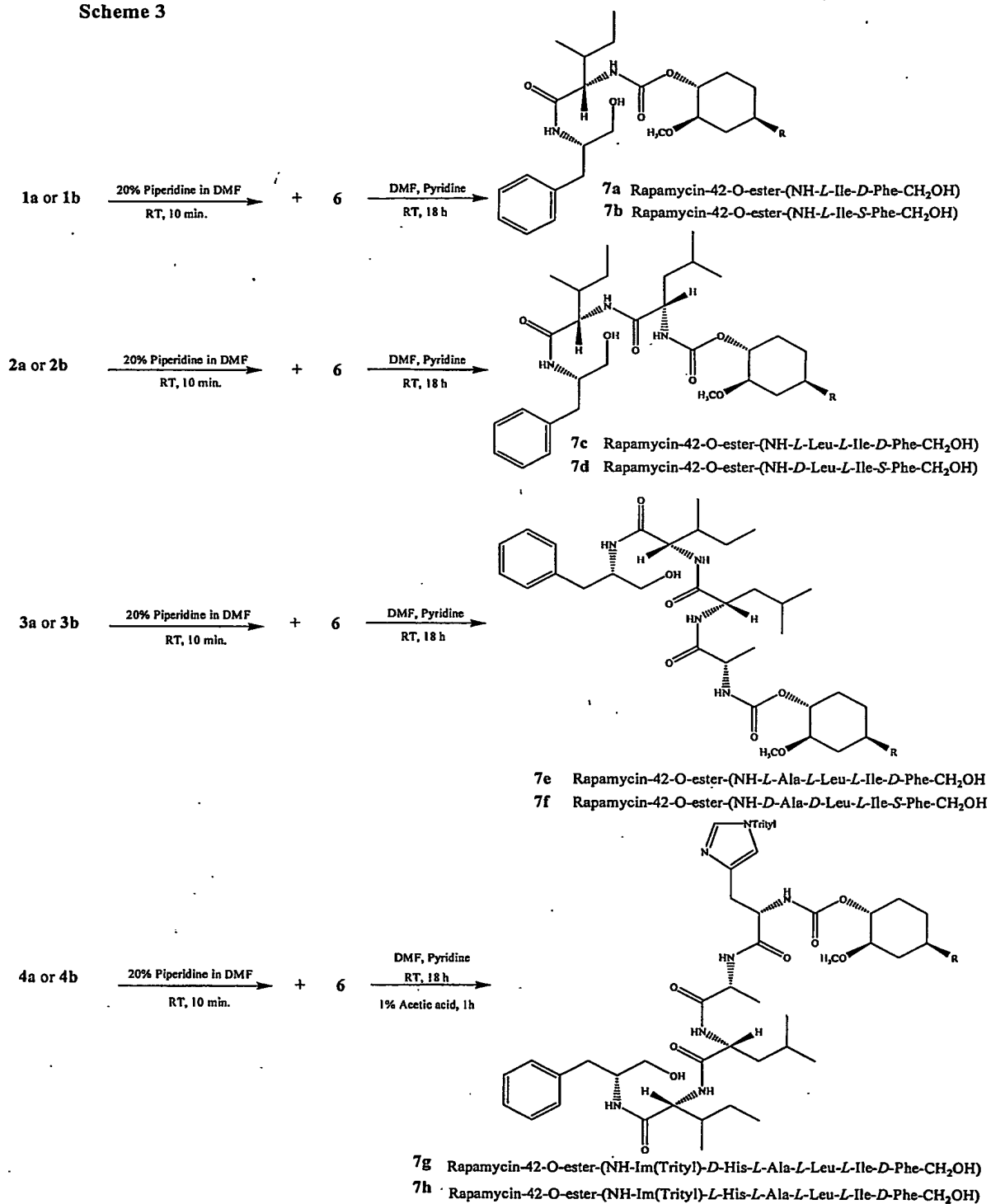
The peptides used to derive 42-O-(4-Nitrophenoxycarbonyl) rapamycin can be synthesized from amino alcohols. The first amino acid is kept as Phe-OH (or 2-amino-3-phenyl-propanol) and performing chain elongation with Fmoc chemistry in solution phase (Scheme 2) using DCC/HOBt as the coupling reagents.

Scheme 2



The subsequent coupling of the peptide sequence with 42-O-(4-Nitrophenoxy carbonyl) rapamycin is done first by de-blocking the Fmoc group under basic conditions (using piperidine for example) and then by coupling the peptide with 42-O-(4-nitrophenoxy carbonyl) rapamycin (6) under basic conditions as shown in **scheme 3** to obtain compounds of general formula 7.

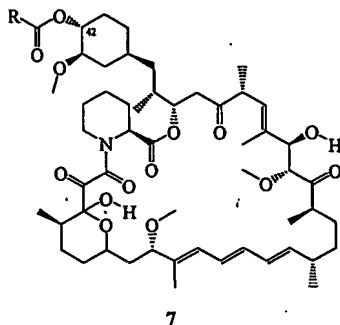
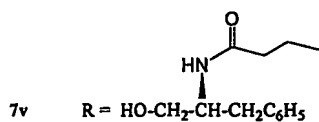
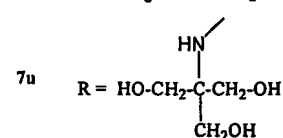
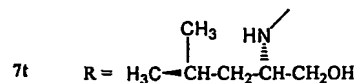
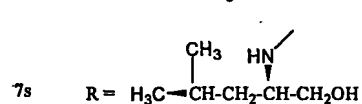
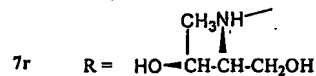
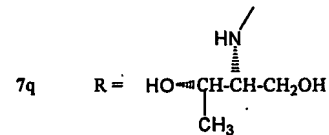
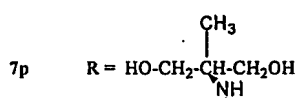
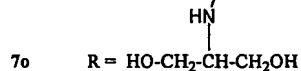
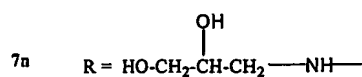
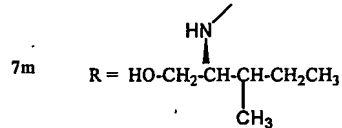
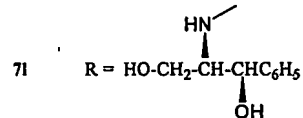
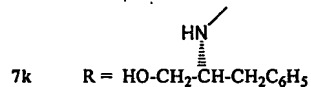
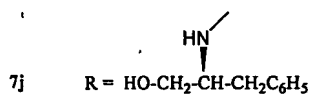
Scheme 3



The derivatives of rapamycin at the 42 position may also be synthesized by conjugating the amino end of amino

alcohols. Compounds 7j to 7v are examples of such amino alcohols-*rapamycin* conjugates (**Sheet 1**).

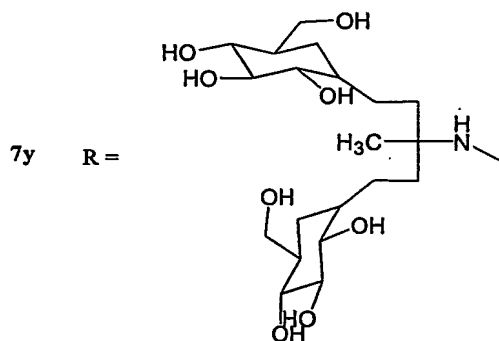
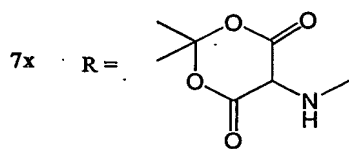
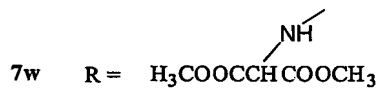
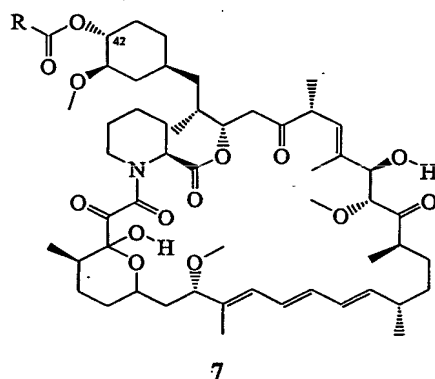
Sheet 1

7a NH-L-Ile-D-Phe-CH<sub>2</sub>OH7b NH-L-Ile-S-Phe-CH<sub>2</sub>OH7c NH-L-Leu-L-Ile-D-Phe-CH<sub>2</sub>OH7d NH-D-Leu-L-Ile-S-Phe-CH<sub>2</sub>OH7e NH-L-Ala-L-Leu-L-Ile-D-Phe-CH<sub>2</sub>OH7f NH-D-Ala-D-Leu-L-Ile-S-Phe-CH<sub>2</sub>OH7g NH-N-Im(Trityl)D-His-L-Ala-L-Leu-L-Ile-D-Phe-CH<sub>2</sub>OH7h NH-N-Im(Trityl)D-His-L-Ala-L-Leu-L-Ile-D-Phe-CH<sub>2</sub>OH7i NH-L-His-L-Ala-L-Lys-L-Arg-L-Arg-L-Leu-L-Ile-D-Phe-CH<sub>2</sub>OH

The conjugations of rapamycin with amino alcohols or peptides comprising an amino alcohol at the "C" terminal of the peptide provides increased hydrophilic character to the compound by virtue of the presence of the free hydroxyl group.

Other rapamycin conjugates (7w, 7x, 7y) exhibiting increase hydrophilicity are shown in sheet 2 below.

Sheet 2



The resulting compounds were screened on a panel of nine human tumor cell lines as listed in **Table 1**.

Quantification of cell proliferation and cell viability was determined by measuring the amount of radioactive [<sup>3</sup>H-methyl]-thymidine incorporated into DNA. The detailed experimental procedure is further described below.

Table 1 Inhibition of H<sup>3</sup> Thymidine uptake (IC<sub>50</sub> in nM)

Compound	A431	Lncap	LS174T	MCF-7	OVCAR-3	SKMEL-2	SK-N-SH	SKOV-3	D341
Adriamycin	50.42	2.544	4.036	3.11	4.773	1.93	0.8959	129.7	7.016
5	341.2	30.33	979.6	0.4077	44.55	1.122	1.329	0.3916	1.561
7a	607.9	153.6	625.6	29.61	415.9	78.06	86.93	80.02	138.2
7b	1838	308.6	1485	53.32	423.6	325.5	143.6	135.5	182.5
7c	286.4	211.8	1698	32.3	260.1	87.37	110.8	72.12	113.6
7d	12748	197.9	NE	52.18	343.1	358.1	137.2	94.35	139.6
7e	489.8	190.6	145.6	66.89	503.1	155.3	90.01	76.58	82.75
7f	NE	NE	NE	NE	NE	NE	NE	NE	NE
7g	NE	NE	3684	2437	NE	945.9	NE	595.1	936.7
7h	2372	374.1	777	85.45	374.9	415.6	222.6	154.7	405.3
7i	28361	NE	NE	1726	1667	NE	NE	13150	7717
7j	505.7	166.3	NE	15.31	263.9	156.6	80.04	55.86	106.4
7k	550.4	145.8	1380	15.94	295.9	109.2	71.32	66.03	259.4
7l	216.8	64.62	23500	6.353	112.9	29.07	15.63	17.15	27.6
7m	625.8	135.9	664	31.31	488.3	147.4	56.69	69.12	86.75
7n	313.5	42.74	55081	2.5333	71.86	11.72	7.271	7.96	14.71

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7o	149.5	39.64	1027	2.579	80.73	48.34	5.837	27.44	36.16
7p	254.3	7.119	2308	1.173	79.33	17.66	7.187	8.797	10.16
7q	312.8	17.37	1631	1.291	56.47	9.154	5.253	6.346	25.21
7r	190.8	21.76	1406	1.86	47.23	8.318	4.839	6.443	5.503
7s	625	189	NE	15.59	355.4	153.9	97.29	93.64	103
7t	277.9	202.4	23978	53.26	366.7	73.15	99.33	64.12	61.17
7u	282.3	13.98	1594	1.539	56.62	6.152	6.402	8.078	20.83
7v	1032	48.34	NE	5.522	95.46	81.01	18.78	10.36	15
7w	NE	83.46	859.3	103.29	NE	378.4	164.11	26.51	NE
7x	NE	3.32	432.8	7.091	NE	112.9	11.3	3.395	NE
7y	NE	573.7	513.6	170	NE	NE	175.1	313.2	NE

As can be seen the compounds can be at least as efficient as rapamycin or in some instances the compounds are more efficient than rapamycin. The *in vitro* efficacy of rapamycin and rapamycin derivatives has been extensively correlated with the *in vivo* efficacy demonstrating that *in vitro* cell proliferation assays are highly predictive of *in vivo* efficacy (see for example Boffa et al. Clin Cancer Res. 2004, 1(10):293-300; Brown et al. PNAS 2003, 100(25):15113-15118; Dudkin et al. Clin Cancer Res. 2001, 7(6):1758-1764).

Without wishing to be bound by theory, the conjugation of peptides and/or amino acids or amino alcohols to rapamycin may provide a "bullet" capable of inactivating the functions of two or more proteins, such as p27<sup>Kip1</sup> and p21<sup>Waf1/Cip1</sup>. This capacity to dual functional inactivation can be advantageous in cases where one of

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the target becomes resistant such as by mutation for example.

Thus the peptide and amino acid derivatives of rapamycin of the present invention are useful for the treatment of conditions in which the control or inhibition of the cell cycle is desirable. Such conditions may comprise but are not limited to: cancer (including solid tumors and leukemia/lymphoma), hyperplasia, psoriasis, fungal infections and the like. It will be appreciated that administration of the compounds of the present invention may be prophylactic to patients susceptible to the above mentioned conditions.

It will also be appreciated that the compounds of the present invention may also be used to treat or prevent hyperproliferative vascular disorders such as restenosis. In particular, the compounds may be applied to, or associated with, surgical stents to prevent restenosis at the site of the stent application in blood vessels. The compounds may for example be incorporated in drug-eluting stents or may be incorporated in a polymer coating applied to the stent. It will be appreciated that the compounds may also be administered to patients already having stents or about to receive such stents. Examples of methods for stent mediated delivery and/or stent coating are described in US patents 6,808,536 and 6,585,764 which are herein incorporated by reference.

When used for restenosis, it is preferred that the compounds of this invention are used to treat restenosis that occurs following an angioplasty procedure.



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The compounds are preferably administered as part of a pharmaceutical composition which may also comprise a pharmaceutically acceptable carrier as would be obvious to one skilled in the art.

5 Preferred routes for the administration of the compounds of the present invention are intravenous, intramuscular, subcutaneous, intraperitoneous, intraarterial, transdermal and oral. Transdermal administrations are understood to include all  
10 administrations across the surface of the body and the inner linings of bodily passages including epithelial and mucosal tissues. Such administrations may be carried out using the present compounds, or pharmaceutically acceptable salts thereof, in lotions, creams, foams,  
15 patches, suspensions, solutions, and suppositories (rectal and vaginal). It will be appreciated that other methods of administration, as would be known to one skilled in the art, may be used such as, for example, local administration at the site of a tumor using a  
20 catheter.

Catheter may be used to guide a carrier containing a compound or formulation of the present invention to be released by the carrier at a desired location. The catheter can be inserted in a lumen of a blood vessel or  
25 of the digestive tract for implanting the carrier. Such catheters are well known in the art.

Solutions or suspensions of these active compounds as a free base or pharmacologically acceptable salt can be prepared in water suitably mixed with a surfactant.

Dispersions can also be prepared in glycerol, liquid polyethylene glycols and mixtures thereof in oils. Under ordinary conditions of storage and use, these preparation contain a preservative to prevent the growth of microorganisms.

The carrier for the injectable form of the compounds of the present invention can be a solvent or dispersion medium containing, for example, water, ethanol, polyol (e.g., glycerol, propylene glycol and liquid polyethylene glycol), suitable mixtures thereof, and vegetable oils.

It will be appreciated that the effective dosage of a rapamycin derivatives of the present invention may vary depending upon the particular compound utilized, the mode of administration, the condition, and severity thereof, of the condition being treated, as well as the various physical factors related to the individual being treated. Satisfactory results may be obtained when the rapamycin derivatives are administered in a daily dosage of about 0.1 µg/kg-100 mg/kg, preferably between 0.001-25 mg/kg, and more preferably between 0.01-5 mg/kg. The effective daily dosages are expected to vary with route of administration.

Oral formulations comprising the compound or composition of the present invention may be contained in conventional oral delivery means including capsules, tablets, and oral liquids and suspensions. The capsules may contain the compound of the present invention with an inert filler or diluent as are well known in the art. Oral formulations herein may utilize delay or time

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release formulations to tailor the pharmacokinetics of the active compound(s) to specific needs.

It is also possible to administer the compounds or composition of the invention directly to the airways in the form of an aerosol.

In another embodiment the compounds may be useful as immunosuppressants and can therefore be useful in treating diseases related to undesired immune responses. Non-limiting example includes preventing graft rejections (host vs graft disease, graft vs host disease), diseases of inflammation and autoimmune diseases such as arthritis.

Patients that are in need of the compounds and composition of the present invention may be identified or diagnosed by a person skilled in the art. For example, cancer diagnosis methods and apparatuses are routinely used and may include blood tests, X-ray radiography and the like.

#### **Screening procedure**

1. Cell culture: For each cell line, culture was carried according to the ATCC Product Information Sheet provided. Cell lines were always freshly thawed prior to each experiment. For all experiments exponentially growing cells were harvested and centrifuged at 1100 rpm, the spent medium was aspirated and cell pellets were resuspended in fresh complete medium. Viable cells were enumerated by trypan blue exclusion using a hemacytometer.

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2. Cells were then seeded in 96 well tissue culture plates in a total volume of 100  $\mu$ L/well. A preliminary experiment should be performed to determine the most appropriate cell density for each individual cell line.

5 Cells were allowed to attach and acclimate overnight.

3. Addition of test compounds: Each test compound was dissolved in DMSO at a final concentration of 2 mM. Each stock compound was then diluted in complete medium (1:100) to obtain a 20  $\mu$ M working solution. The working

10 solution was used for further serial dilutions to obtain concentrations of 200 nM, 20 nM, 2 nM, .2 nM and .02 nM. 100  $\mu$ L of each dilution was added to the 100  $\mu$ L cell cultures (3 replicates), to give final test concentrations of 100 nM, 10 nM, 1 nM, .1 nM and .01 nM.

15 Using this system of dilutions, the maximum concentration of DMSO to which the cells were exposed was 0.01% v/v. Therefore, 0.01% DMSO was added to control cells to which no test compounds was added. On each plate two positive controls were included: Adriamycin Hydrochloride and

20 Rapamycin. For each positive control 3 concentrations were chosen in a range that achieved an LC<sub>50</sub>, this range is cell line specific and must be predetermined in a pilot experiment. Plates were incubated for 96 hours prior to harvesting. [<sup>3</sup>H-methyl]-thymidine incorporation:

25 After 80 hours incubation, 10  $\mu$ L (0.5  $\mu$ Ci) of [<sup>3</sup>H-methyl]-thymidine diluted in 1X HBSS was added and plates were incubated overnight.

Growth medium was removed from each well and 100  $\mu$ L of Trypsin-EDTA was added. The plate was incubated at

30 37°C until cells were trypsinized (check under

microscope). The detached cells were harvested using a semiautomatic cell harvester. Filters were dried prior to addition to scintillation vials. two mL of scintillation fluid was automatically dispensed into each  
5 vial and counting was effected on program 1 ( $^3\text{H}$ , 1 min, DPM).

The average and standard error from the DPM counts of replicate samples were calculated. The  $\text{IC}_{50}$  values of these screening results are listed in **Table 1**.

#### 10 EXAMPLE 1

##### **Synthesis of Fmoc-NH-L-Ile-D-Phe-CH<sub>2</sub>OH (1a)**

(R)(+)-2-amino-3-phenyl-1-propanol (427 mg, 2.83 mmol) dissolved in dry DMF (20 mL), stirred under nitrogen, to this stirred mixture at 25°C DCC (699 mg,  
15 3.39 mmol), HOBt (457 mg, 3.38 mmol) was added with constant stirring. After 10 minutes of stirring N-(9-fluorenylmethoxycarbonyl)-L-isoleucine (1.0g, 2.83 mmol) was added to the above mixture and then stirred at 25°C for 14 h. After 14 h white colored crystals precipitated  
20 out (DCU), which was filtered. The filtrate was checked on TLC (2% MeOH: CH<sub>2</sub>Cl<sub>2</sub>) which showed formation of a new compound at higher  $R_f$  (0.6), LC/MS also showed molecular ion peak corresponding to the dipeptide **1a** with other impurities peaks. The crude product was chromatographed  
25 on silica gel (Silica gel 60, 63-200 $\mu$ ) column and eluted with 2% MeOH: CH<sub>2</sub>Cl<sub>2</sub> to give 650 mg of the dipeptide Fmoc-NH-L-Ile-D-Phe-CH<sub>2</sub>OH (**1a**) as a white solid. Checked on LC/MS which showed  $M^+ + 1$  (487.4) and  $M^+ + \text{Na}$  (509.2).

EXAMPLE 2**Synthesis of Fmoc-NH-L-Ile-S-Phe-CH<sub>2</sub>OH (1b)**

S) (+)-2-amino-3-phenyl-1-propanol (1.3g mg, 8.59 mmol) dissolved in dry DMF (100 mL) , stirred under nitrogen, to this stirred mixture at 25°C DCC (1.94g, 10.9 mmol), HOBt (1.27g, 10.9 mmol) was added with constant stirring. After 10 minutes of stirring N-(9-fluorenylmethoxycarbonyl)-L-isoleucine (3.0g, 2.83 mmol) was added to the above mixture and then stirred at 25°C for 14 h. After 14 h white colored crystals precipitated out (DCU), which was filtered. The filtrate was checked on TLC (2% MeOH: CH<sub>2</sub>Cl<sub>2</sub>) which showed formation of a new compound at higher R<sub>f</sub> (0.6), LC/MS also showed molecular ion peak corresponding to the dipeptide **1b** with other impurities peaks. The crude product was chromatographed on silica gel (Silica gel 60, 63-200μ) column and eluted with 2% MeOH: CH<sub>2</sub>Cl<sub>2</sub> to give 2.8g of the dipeptide Fmoc-NH-L-Ile-S-Phe-CH<sub>2</sub>OH (**1b**) as a white solid. Checked on LC/MS which showed M<sup>+</sup>+1 (487.2) and M<sup>+</sup>+ Na (509.2).

EXAMPLE 3**Synthesis of Fmoc-NH-L-Leu-L-Ile-D-Phe-CH<sub>2</sub>OH (2a)**

Dipeptide Fmoc-NH-L-Ile-D-Phe-CH<sub>2</sub>OH (**1a**) (520 mg, 1.06 mmol) was taken in 20% piperidine in DMF (2 mL) and stirred for 15 minutes at 25°C, TLC examination showed complete removal of the Fmoc protecting group (NH<sub>2</sub>-L-Ile-D-Phe-CH<sub>2</sub>OH), further confirmed by LC/MS examination, which showed M<sup>+</sup>-1 (263.3) peak. The reaction mixture was concentrated in vacuo, excess of the piperidine was

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removed by co-evaporating with toluene (2x10 mL), the free amino dipeptide was further dried over high vacuum for 30 minutes, and then redissolved in dry DMF (2 mL) and added to the mixture of N-(9-fluorenylmethoxycarbonyl)-L-leucine (38.1 mg, 1.06 mmol), DCC (262.0 mg, 1.27 mmol) and HOBt (171.4 mg, 1.27 mmol) in dry DMF (20 mL) at 25°C. The stirring was further continued for 14 h at 25°C. After 14 h white colored crystals precipitated out (DCU), which was filtered. The filterate was checked on TLC (3% MeOH: CH<sub>2</sub>Cl<sub>2</sub>) which showed formation of a new compound at higher R<sub>f</sub> (0.5), LC/MS also showed molecular ion peak corresponding to the tripeptide **2a** with other impurities peaks. The crude product was chromatographed on silica gel (Silica gel 60, 63-200μ) column and eluted with 3% MeOH: CH<sub>2</sub>Cl<sub>2</sub> to give 310 mg of the tripeptide Fmoc-NH-L-Leu-L-Ile-D-Phe-CH<sub>2</sub>OH (**2a**) as a white solid. Checked on LC/MS which showed M<sup>+</sup>+1 (600.3) and M<sup>+</sup>+ Na (622.3).

#### EXAMPLE 4

##### **Synthesis of Fmoc-NH-D-Leu-L-Ile-S-Phe-CH<sub>2</sub>OH (2b)**

Dipeptide Fmoc-NH-L-Ile-S-Phe-CH<sub>2</sub>OH (**1b**) (2.0g, 4.11 mmol) was taken in 20% piperidine in DMF (20 mL) and stirred for 15 minutes at 25°C, TLC examination showed complete removal of the Fmoc protecting group (NH<sub>2</sub>-L-Ile-S-Phe-CH<sub>2</sub>OH), further confirmed by LC/MS examination, which showed M<sup>+</sup>-1 (263.3) peak. The reaction mixture was concentrated in vacuo, excess of the piperidine was removed by co-evaporating with toluene (2x20 mL), the free amino dipeptide was further dried over high vacuum

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for 30 minutes, and then redissolved in dry DMF (5 mL) and added to the mixture of N-(9-fluorenylmethoxycarbonyl)-*D*-leucine (1.59g, 4.52 mmol), DCC (931 mg, 4.52 mmol) and HOBt (610.0 mg, 4.52 mmol) in dry DMF (150 mL) at 25°C. The stirring was further continued for 14 h at 25°C. After 14 h white colored crystals precipitated out (DCU), which was filtered. The filtrate was checked on TLC (5% MeOH: CH<sub>2</sub>Cl<sub>2</sub>) which showed formation of a new compound at higher R<sub>f</sub> (0.5), LC/MS also showed molecular ion peak corresponding to the tripeptide **2b** with other impurities peaks. The crude product was chromatographed on silica gel (Silica gel 60, 63-200μ) column and eluted with 4% MeOH: CH<sub>2</sub>Cl<sub>2</sub> to give 450 mg of the tripeptide Fmoc-NH-*D*-Leu-*L*-Ile-*S*-Phe-CH<sub>2</sub>OH (**2b**) as a white solid. Checked on LC/MS which showed M<sup>+</sup>+1 (600.3) and M<sup>+</sup>+ Na (622.3).

#### EXAMPLE 5

##### **Synthesis of Fmoc-NH-*L*-Ala-*L*-Leu-*L*-Ile-*D*-Phe-CH<sub>2</sub>OH (3a)**

Tripeptide Fmoc-NH-*L*-Leu-*L*-Ile-*D*-Phe-CH<sub>2</sub>OH (**2a**) (75 mg, 0.125 mmol) was taken in 20% piperidine in DMF (0.5 mL) and stirred for 15 minutes at 25°C, TLC examination showed complete removal of the Fmoc protecting group (NH<sub>2</sub>-*L*-Leu-*L*-Ile-*D*-Phe-CH<sub>2</sub>OH), further confirmed by LC/MS examination, which showed M<sup>+</sup>-1 (376.1) peak. The reaction mixture was concentrated in vacuo, excess of the piperidine was removed by co-evaporating with toluene (2x2 mL), the free amino tripeptide was further dried over high vacuum for 30 minutes, and then redissolved in dry DMF (0.5 mL) and added to the mixture of N-(9-



fluorenylmethoxycarbonyl)-L-alanine (42.0 mg, 0.137 mmol), DCC (28.0 mg, 0.137 mmol) and HOBt (18.4 mg, 0.137 mmol) in dry DMF (2.5 mL) at 25°C. The stirring was further continued for 14 h at 25°C. After 14 h white colored crystals precipitated out (DCU), which was filtered. The filtrate was checked on TLC (10% MeOH: CH<sub>2</sub>Cl<sub>2</sub>) which showed formation of a new compound at higher R<sub>f</sub> (0.45), LC/MS also showed molecular ion peak corresponding to the tetrapeptide **3a** with other impurities peaks. The crude product was chromatographed on silica gel (Silica gel 60, 63-200μ) column and eluted with 8% MeOH: CH<sub>2</sub>Cl<sub>2</sub> to give 70 mg of the tetrapeptide Fmoc-NH-L-Ala-L-Leu-L-Ile-D-Phe-CH<sub>2</sub>OH (**3a**) as a white solid. Checked on LC/MS which showed M<sup>+</sup>+1 (671.3) and M<sup>+</sup>+Na (693.3).

#### EXAMPLE 6

##### **Synthesis of Fmoc-NH-D-Ala-D-Leu-L-Ile-S-Phe-CH<sub>2</sub>OH (**3b**)**

Tripeptide Fmoc-NH-D-Leu-D-Ile-S-Phe-CH<sub>2</sub>OH (**2b**) (330 mg, 0.550 mmol) was taken in 20% piperidine in DMF (1.5 mL) and stirred for 15 minutes at 25°C, TLC examination showed complete removal of the Fmoc protecting group (NH<sub>2</sub>-D-Leu-D-Ile-S-Phe-CH<sub>2</sub>OH), further confirmed by LC/MS examination, which showed M-1 (376.1) peak. The reaction mixture was concentrated in vacuo, excess of the piperidine was removed by co-evaporating with toluene (2x5 mL), the free amino tripeptide was further dried over high vacuum for 30 minutes, and then redissolved in dry DMF (1.0 mL) and added to the mixture of N-(9-fluorenylmethoxycarbonyl)-D-alanine (188.4 mg, 0.606

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mmol), DCC (124.0 mg, 0.606 mmol) and HOBt (81 mg, 0.606 mmol) in dry DMF (2.5 mL) at 25°C. The stirring was further continued for 14 h at 25°C. After 14 h white colored crystals precipitated out (DCU), which was  
5 filtered. The filtrate was checked on TLC (10% MeOH: CH<sub>2</sub>Cl<sub>2</sub>) which showed formation of a new compound at higher R<sub>f</sub> (0.40), LC/MS also showed molecular ion peak corresponding to the tetrapeptide **3b** with other impurities peaks. The crude product was chromatographed  
10 on silica gel (Silica gel 60, 63-200μ) column and eluted with 7% MeOH: CH<sub>2</sub>Cl<sub>2</sub> to give 40 mg of the tetrapeptide Fmoc-NH-D-Ala-D-Leu-D-Ile-S-Phe-CH<sub>2</sub>OH (**3b**) as a white solid. Checked on LC/MS which showed M<sup>+</sup>+1 (671.4) and M<sup>+</sup>+Na (693.4).

#### 15 EXAMPLE 7

##### **Synthesis of N-α-Fmoc-N-Im(trityl)-D-His-L-Ala-L-Leu-L-Ile-D-Phe-CH<sub>2</sub>OH (4a)**

Tetrapeptide Fmoc-NH-L-Ala-L-Leu-L-Ile-D-Phe-CH<sub>2</sub>OH (**3a**) (300 mg, 0.447 mmol) was taken in 20% piperidine in  
20 DMF (2.5 mL) and stirred for 15 minutes at 25°C, TLC examination showed complete removal of the Fmoc protecting group (NH<sub>2</sub>-L-Ala-L-Leu-L-Ile-D-Phe-CH<sub>2</sub>OH), further confirmed by LC/MS examination, which showed M<sup>+</sup>+1 (450.3) peak. The reaction mixture was concentrated in  
25 vacuo, excess of the piperidine was removed by co-evaporating with toluene (2x5 mL), the free amino tetrapeptide was further dried over high vacuum for 30 minutes, and then redissolved in dry DMF (1.5 mL) and added to the mixture of N- α - (9-

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fluorenylmethoxycarbonyl)-N-Im(trityl)-D-histidine (304.8 mg, 0.492 mmol), DCC (101.3 mg, 0.492 mmol) and HOBT (66.4 mg, 0.492 mmol) in dry DMF (2.5 mL) at 25°C. The stirring was further continued for 14 h at 25°C. After 14 h white colored crystals precipitated out (DCU), which was filtered. The filtrate was checked on TLC (15% MeOH: CH<sub>2</sub>Cl<sub>2</sub>) which showed formation of a new compound at higher R<sub>f</sub> (0.55), LC/MS also showed molecular ion peak corresponding to the pentapeptide **4a** with other impurities peaks. The crude product was chromatographed on silica gel (Silica gel 60, 63-200μ) column and eluted with 10% MeOH: CH<sub>2</sub>Cl<sub>2</sub> to give 376 mg of the pentapeptide N-α-Fmoc-N-Im(trityl)-D-His-L-Ala-L-Leu-L-Ile-D-Phe-CH<sub>2</sub>OH (**4a**) as white solid. Checked on LC/MS which showed M<sup>+</sup>+1 (1050.6).

#### EXAMPLE 8

##### **Synthesis of N-α-Fmoc-N-Im(trityl)-L-His-L-Ala-L-Leu-L-Ile-D-Phe-CH<sub>2</sub>OH (**4b**)**

Tetrapeptide Fmoc-NH-L-Ala-L-Leu-L-Ile-D-Phe-CH<sub>2</sub>OH (**3a**) (300 mg, 0.447 mmol) was taken in 20% piperidine in DMF (2.5 mL) and stirred for 15 minutes at 25°C, TLC examination showed complete removal of the Fmoc protecting group (NH<sub>2</sub>-L-Ala-L-Leu-L-Ile-D-Phe-CH<sub>2</sub>OH), further confirmed by LC/MS examination, which showed M<sup>+</sup>+1 (450.3) peak. The reaction mixture was concentrated in vacuo, excess of the piperidine was removed by co-evaporating with toluene (2x5 mL), the free amino tetrapeptide was further dried over high vacuum for 30 minutes, and then redissolved in dry DMF (1.5 mL) and

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added to the mixture of N-  $\alpha$  -(9-fluorenylmethoxycarbonyl)-N-Im(trityl)-L-histidine (304.8 mg, 0.492 mmol), DCC (101.3 mg, 0.492 mmol) and HOBT (66.4 mg, 0.492 mmol) in dry DMF (2.5 mL) at 25°C. The stirring was further continued for 14 h at 25°C. After 14 h white colored crystals precipitated out (DCU), which was filtered. The filtrate was checked on TLC (15% MeOH: CH<sub>2</sub>Cl<sub>2</sub>) which showed formation of a new compound at higher R<sub>f</sub> (0.55), LC/MS also showed molecular ion peak corresponding to the pentapeptide **4b** with other impurities peaks. The crude product was chromatographed on silica gel (Silica gel 60, 63-200 $\mu$ ) column and eluted with 10% MeOH: CH<sub>2</sub>Cl<sub>2</sub> to give 376 mg of the pentapeptide N- $\alpha$ -Fmoc-N-Im(trityl)-L-His-L-Ala-L-Leu-L-Ile-D-Phe-CH<sub>2</sub>OH (**4b**) as white solid. Checked on LC/MS which showed M<sup>+</sup>+1 (1050.6).

#### EXAMPLE 9

##### **42-O-(4-Nitrophenoxy carbonyl) rapamycin (6)**

To a solution of 5.00 g (5.47 mmol) of rapamycin (**5**) in 40 ml of dichloromethane cooled at -78°C with dry ice and acetone bath was added 650  $\mu$ l of dry pyridine and 1.65 g of p-nitrophenyl chloroformate dissolved in 10 ml of dichloromethane. The reaction mixture was allowed to warm to ambient temperature and stirred for two hours under nitrogen. After two hours 325  $\mu$ l of dry pyridine and 555 mg of the p-nitrophenyl chloroformate was added to the above reaction mixture. The reaction mixture was stirred under nitrogen for 18h. The progress of the reaction was monitored by mass spectrum. After 18h the

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reaction mixture was concentrated in vacuum and partitioned between ether and water. The organic phase was washed with 0.1N HCl (3x 400ml) than with saturated brine solution (2x 100ml), dried over sodium sulphate, 5 filtered and concentrated in vacuum to give the pale yellow solid, which was purified on silica gel (Silica gel 60, 63-200 $\mu$ ). Elution with 40% and then 50% ethyl acetate: Hexane gave 4.7 g of the title compound (6) as yellow solid.

10  $^1\text{H}$  NMR ( $\text{CDCl}_3$ ):  $\delta$  8.27 and 7.39 (aromatic-H, 4H), 4.63 (42C, 1H): Mass spectra: Positive M + Na 1101.5 (100%): Negative M-1 1077.5 (100%).

#### EXAMPLE 10

##### **Synthesis of Rapamycin- 42-O-ester-(NH-L-Ile-D-Phe-CH<sub>2</sub>OH)**

15 (7a)

Dipeptide Fmoc-NH-L-Ile-D-Phe-CH<sub>2</sub>OH (1a) (67.0 mg, 0.138 mmol) was taken in 20% piperidine in DMF (0.5 mL) and stirred for 15 minutes at 25°C, TLC examination showed complete removal of the Fmoc protecting group (NH<sub>2</sub>-L-Ile-D-Phe-CH<sub>2</sub>OH), further confirmed by LC/MS examination, 20 which showed M<sup>+</sup>-1 (263.3) peak. The reaction mixture was concentrated in vacuo, excess of the piperidine was removed by co-evaporating with toluene (2x2.5 mL), the free amino dipeptide was further dried over high vacuum 25 for 30 minutes, and then redissolved in dry DMF (2 mL) and added to the mixture of 42-O-(4-Nitrophenoxy carbonyl)rapamycin (6) (100 mg, 0.092 mmol) and pyridine (50  $\mu\text{L}$ ) in dry DMF (10 mL) at 25°C. The

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stirring was further continued for 14 h at 25°C. After 14 h reaction mixture was checked on TLC (pure ethyl acetate) which showed formation of a new compound at lower  $R_f$  (0.5) than the starting material, LC/MS also showed molecular ion peak corresponding to the conjugated product **7a** with other impurities peaks. The crude product was chromatographed on silica gel (Silica gel 60, 63-200 $\mu$ ) column and eluted with pure ethyl acetate to afford 50 mg (45% yield) of the conjugate Rapamycin-42-O-ester-(NH-*L*-Ile-*D*-Phe-CH<sub>2</sub>OH) **7a** as a light yellow colored solid. Checked on LC/MS which showed M-1 1202.7 (100%).

#### EXAMPLE 11

#### **Synthesis of Rapamycin- 42-O-ester-(NH-*L*-Ile-*S*-Phe-CH<sub>2</sub>OH) (**7b**)**

Dipeptide Fmoc-NH-*L*-Ile-*S*-Phe-CH<sub>2</sub>OH (**1b**) (67.0 mg, 0.138 mmol) was taken in 20% piperidine in DMF (0.2 mL) and stirred for 15 minutes at 25°C, TLC examination showed complete removal of the Fmoc protecting group (NH<sub>2</sub>-*L*-Ile-*S*-Phe-CH<sub>2</sub>OH), further confirmed by LC/MS examination, which showed M<sup>+</sup>-1 (263.3) peak. The reaction mixture was concentrated in vacuo, excess of the piperidine was removed by co-evaporating with toluene (2x2.5 mL), the free amino dipeptide was further dried over high vacuum for 30 minutes, and then redissolved in dry DMF (2 mL) and added to the mixture of 42-O-(4-Nitrophenoxy carbonyl)rapamycin (**6**) (100 mg, 0.092 mmol) and pyridine (50  $\mu$ L) in dry DMF (10 mL) at 25°C. The stirring was further continued for 14 h at 25°C. After 14 h reaction mixture was checked on TLC (pure ethyl

acetate) which showed formation of a new compound at lower  $R_f$  (0.4) than the starting material, LC/MS also showed molecular ion peak corresponding to the conjugated product **7b** with other impurities peaks. The crude product was chromatographed on silica gel (Silica gel 60, 63-200 $\mu$ ) column and eluted with pure ethyl acetate to afford 18.8 mg (20% yield) of the conjugate Rapamycin-42-O-ester-(NH-*L*-Ile-*S*-Phe-CH<sub>2</sub>OH) **7b** as a light yellow colored solid. Checked on LC/MS which showed  $M-1$  1202.6 (100%).

#### 10 EXAMPLE 12

##### **Synthesis of Rapamycin- 42-O-ester-(NH-*L*-Leu-*L*-Ile-*D*-Phe-CH<sub>2</sub>OH) (**7c**)**

Dipeptide Fmoc-NH-*L*-Leu-*L*-Ile-*D*-Phe-CH<sub>2</sub>OH (**2a**) (60.6 mg, 0.101 mmol) was taken in 20% piperidine in DMF (0.3 mL) and stirred for 15 minutes at 25°C, TLC examination showed complete removal of the Fmoc protecting group (NH<sub>2</sub>-*L*-Leu-*L*-Ile-*D*-Phe-CH<sub>2</sub>OH), further confirmed by LC/MS examination, which showed  $M^+-1$  (376.1) peak. The reaction mixture was concentrated in vacuo, excess of the piperidine was removed by co-evaporating with toluene (2x2.5 mL), the free amino tripeptide was further dried over high vacuum for 30 minutes, and then redissolved in dry DMF (2 mL) and added to the mixture of 42-O-(4-Nitrophenoxy carbonyl)rapamycin (**6**) (100 mg, 0.092 mmol) and pyridine (50  $\mu$ L) in dry DMF (10 mL) at 25°C. The stirring was further continued for 14 h at 25°C. After 14 h reaction mixture was checked on TLC (pure ethyl acetate) which showed formation of a new compound at lower  $R_f$  (0.5) than the starting material, LC/MS also

showed molecular ion peak corresponding to the conjugated product **7c** with other impurities peaks. The crude product was chromatographed on silica gel (Silica gel 60, 63-200 $\mu$ ) column and eluted with pure ethyl acetate to afford  
5 45 mg (37% yield) of the conjugate Rapamycin-42-O-ester-(NH-*L*-Leu-*L*-Ile-*D*-Phe-CH<sub>2</sub>OH) **7c** as a white solid. Checked on LC/MS which showed M-1 1315.6 (100%).

### EXAMPLE 13

#### **Synthesis of Rapamycin- 42-O-ester-(NH-*D*-Leu-*L*-Ile-*S*-Phe-CH<sub>2</sub>OH) (**7d**)** 10

Dipeptide Fmoc-NH-*D*-Leu-*L*-Ile-*S*-Phe-CH<sub>2</sub>OH (**2b**) (60.6 mg, 0.101 mmol) was taken in 20% piperidine in DMF (1.0 mL) and stirred for 15 minutes at 25°C, TLC examination showed complete removal of the Fmoc protecting group (NH<sub>2</sub>-*D*-Leu-*L*-Ile-*S*-Phe-CH<sub>2</sub>OH), further confirmed by LC/MS  
15 examination, which showed M<sup>+</sup>-1 (376.1) peak. The reaction mixture was concentrated in vacuo, excess of the piperidine was removed by co-evaporating with toluene (2x5.0 mL), the free amino tripeptide was further dried  
20 over high vacuum for 30 minutes, and then redissolved in dry DMF (2 mL) and added to the mixture of 42-O-(4-Nitrophenoxy carbonyl) rapamycin (**6**) (100 mg, 0.092 mmol) and pyridine (50  $\mu$ L) in dry DMF (10 mL) at 25°C. The stirring was further continued for 14 h at 25°C. After 14  
25 h reaction mixture was checked on TLC (pure ethyl acetate) which showed formation of a new compound at lower R<sub>f</sub> (0.45) than the starting material, LC/MS also showed molecular ion peak corresponding to the conjugated product **7d** with other impurities peaks. The crude product



was chromatographed on silica gel (Silica gel 60, 63-200 $\mu$ ) column and eluted with pure ethyl acetate to afford 44 mg (36% yield) of the conjugate Rapamycin-42-O-ester-(NH-*D*-Leu-*L*-Ile-*S*-Phe-CH<sub>2</sub>OH) **7d** as a white solid. Checked  
5 on LC/MS which showed M-1 1315.6 (100%).

#### EXAMPLE 14

#### **Synthesis of Rapamycin- 42-O-ester-(NH- *L*-Ala-*L*-Leu-*L*-Ile-*D*-Phe-CH<sub>2</sub>OH) (**7e**)**

Tetrapeptide Fmoc-NH-*L*-Ala-*L*-Leu-*L*-Ile-*D*-Phe-CH<sub>2</sub>OH  
10 (**3a**) (60 mg, 0.089 mmol) was taken in 20% piperidine in DMF (1.5 mL) and stirred for 15 minutes at 25°C, TLC examination showed complete removal of the Fmoc protecting group (NH<sub>2</sub>-*L*-Ala-*L*-Leu-*L*-Ile-*D*-Phe-CH<sub>2</sub>OH), further confirmed by LC/MS examination, which showed M<sup>+</sup>+1  
15 (450.3) peak. The reaction mixture was concentrated in vacuo, excess of the piperidine was removed by co-evaporating with toluene (2x2.5 mL), the free amino tetrapeptide was further dried over high vacuum for 30 minutes, and then redissolved in dry DMF (1.5 mL) and  
20 added to the mixture of 42-O-(4-Nitrophenoxy carbonyl) rapamycin (**6**) (100 mg, 0.092 mmol) and pyridine (50  $\mu$ L) in dry DMF (10 mL) at 25°C. The stirring was further continued for 14 h at 25°C. After 14 h reaction mixture was checked on TLC (5% MeOH:CH<sub>2</sub>Cl<sub>2</sub>) which showed formation  
25 of a new compound at lower R<sub>f</sub> (0.45) than the starting material, LC/MS also showed molecular ion peak corresponding to the conjugated product **7e** with other impurities peaks. The crude product was purified on preparative TLC using 5% MeOH: CH<sub>2</sub>Cl<sub>2</sub> as the developing

solvent system to afford 9.8mg (10% yield) of the conjugate Rapamycin-42-O-ester-(NH- L-Ala-L-Leu-L-Ile-D-Phe-CH<sub>2</sub>OH) **7e** as a white solid. Checked on LC/MS which showed M<sup>+</sup>+Na 1410.8 (100%).

5 EXAMPLE 15

**Synthesis of Rapamycin- 42-O-ester-(NH- D-Ala-D-Leu-L-Ile-S-Phe-CH<sub>2</sub>OH) (7f)**

Tetrapeptide Fmoc-NH-D-Ala-D-Leu-L-Ile-S-Phe-CH<sub>2</sub>OH (**3b**) (60 mg, 0.089 mmol) was taken in 20% piperidine in  
10 DMF (1.5 mL) and stirred for 15 minutes at 25°C, TLC examination showed complete removal of the Fmoc protecting group (NH<sub>2</sub>-D-Ala-D-Leu-L-Ile-S-Phe-CH<sub>2</sub>OH), further confirmed by LC/MS examination, which showed M<sup>+</sup>+1 (450.3) peak. The reaction mixture was concentrated in  
15 vacuo, excess of the piperidine was removed by co-evaporating with toluene (2x2.5 mL), the free amino tetrapeptide was further dried over high vacuum for 30 minutes, and then redissolved in dry DMF (1.5 mL) and added to the mixture of 42-O-(4-Nitrophenoxy carbonyl)  
20 rapamycin (**6**) (100 mg, 0.092 mmol) and pyridine (50 µL) in dry DMF (10 mL) at 25°C. The stirring was further continued for 14 h at 25°C. After 14 h reaction mixture was checked on TLC (5% MeOH:CH<sub>2</sub>Cl<sub>2</sub>) which showed formation of a new compound at lower R<sub>f</sub> (0.45) than the starting  
25 material, LC/MS also showed molecular ion peak corresponding to the conjugated product **7f** with other impurities peaks. The crude product was purified on preparative TLC using 5% MeOH: CH<sub>2</sub>Cl<sub>2</sub> as the developing solvent system to afford 12.3 mg (11%yield) of the

conjugate Rapamycin-42-O-ester-(NH- *D*-Ala-*D*-Leu-*L*-Ile-*S*-Phe-CH<sub>2</sub>OH) **7f** as a white solid. Checked on LC/MS which showed  $M^+ + Na$  1410.8 (100%).

#### EXAMPLE 16

#### 5 **Synthesis of Rapamycin- 42-O-ester-(NH-N-Im(Trityl)-*D*-His-*L*-Ala-*L*-Leu-*L*-Ile-*D*-Phe-CH<sub>2</sub>OH (7g)**

Pentapeptide N- $\alpha$ -Fmoc-N-Im(trityl)*D*-His-*L*-Ala-*L*-Leu-*L*-Ile-*D*-Phe-CH<sub>2</sub>OH (**4**) (150 mg, 0.142 mmol) was taken in 20% piperidine in DMF (2.5 mL) and stirred for 15 minutes  
10 at 25°C, TLC examination showed complete removal of the Fmoc protecting group (NH<sub>2</sub>-N-Im(trityl)-*D*-His-*L*-Ala-*L*-Leu-*L*-Ile-*D*-Phe-CH<sub>2</sub>OH), further confirmed by LC/MS examination, which showed  $M^+ + 1$  (828) peak. The reaction mixture was concentrated in vacuo, excess of the  
15 piperidine was removed by co-evaporating with toluene (2x5 mL), the free amino pentapeptide was further dried over high vacuum for 30 minutes, and then redissolved in dry DMF (1.5 mL) and added to the mixture of 42-O-(4-Nitrophenoxy carbonyl) rapamycin (**6**) (158 mg, 0.147 mmol)  
20 and pyridine (50  $\mu$ L) in dry DMF (10 mL) at 25°C. The stirring was further continued for 14 h at 25°C. After 14 h reaction mixture was checked on TLC (5% MeOH:CH<sub>2</sub>Cl<sub>2</sub>) which showed formation of a new compound at lower  $R_f$  (0.45) then the starting material, LC/MS also showed  
25 molecular ion peak corresponding to the conjugated product **7g** with other impurities peaks. The crude product was chromatographed on silica gel (Silica gel 60, 63-200 $\mu$ ) column and eluted with pure 10% MeOH: CH<sub>2</sub>Cl<sub>2</sub> to afford 41 mg (16% yield) of the conjugate Rapamycin-42-O-

ester-(NH-N-Im(Trityl))-D-His-L-Ala-L-Leu-L-Ile-D-Phe-CH<sub>2</sub>OH (7g) as a white solid. Checked on LC/MS which showed M-1 1765.6 (90%) and 1766.8 (100%).

#### EXAMPLE 17

#### 5 **Synthesis of Rapamycin- 42-O-ester-(NH-N-Im(Trityl))-L-His-L-Ala-L-Leu-L-Ile-D-Phe-CH<sub>2</sub>OH (7h)**

Pentapeptide N- $\alpha$ -Fmoc-N-Im(trityl)-L-His-L-Ala-L-Leu-L-Ile-D-Phe-CH<sub>2</sub>OH (4b) (150 mg, 0.142 mmol) was taken in 20% piperidine in DMF (2.5 mL) and stirred for 15 minutes at 25°C, TLC examination showed complete removal of the Fmoc protecting group (NH<sub>2</sub>-N-Im(trityl)-D-His-L-Ala-L-Leu-L-Ile-D-Phe-CH<sub>2</sub>OH), further confirmed by LC/MS examination, which showed M<sup>+</sup>+1 (828) peak. The reaction mixture was concentrated in vacuo, excess of the piperidine was removed by co-evaporating with toluene (2x5 mL), the free amino pentapeptide was further dried over high vacuum for 30 minutes, and then redissolved in dry DMF (1.5 mL) and added to the mixture of 42-O-(4-Nitrophenoxy carbonyl) rapamycin (6) (158 mg, 0.147 mmol) and pyridine (50  $\mu$ L) in dry DMF (10 mL) at 25°C. The stirring was further continued for 14 h at 25°C. After 14 h reaction mixture was checked on TLC (5% MeOH:CH<sub>2</sub>Cl<sub>2</sub>) which showed formation of a new compound at lower R<sub>f</sub> (0.45) than the starting material, LC/MS also showed molecular ion peak corresponding to the conjugated product 7h with other impurities peaks. The crude product was chromatographed on silica gel (Silica gel 60, 63-200 $\mu$ ) column and eluted with pure 10% MeOH: CH<sub>2</sub>Cl<sub>2</sub> to afford 41 mg (16% yield) of the conjugate Rapamycin-42-O-

ester-(NH-N-Im(Trityl)-L-His-L-Ala-L-Leu-L-Ile-D-Phe-CH<sub>2</sub>OH (7h) as a white solid. Checked on LC/MS which showed M-1 1765.6 (90%) and 1766.8 (100%).

#### EXAMPLE 18

#### 5 **Synthesis of NH<sub>2</sub>-N-Im(Trityl)-L-His-L-Ala-L-Lys-L-Arg-L-Arg-L-Leu-L-Ile-D-Phe-CH<sub>2</sub>OH (7i)**

Octapeptide (7i) was synthesized by the reported procedure (Atkinson, G.E et al. Bioorganic Med. Chem. Lett. 2002, 12, 2501-2505) using solid phase method by  
10 solid phase method on a peptide synthesizer., LCMS 1039.5379 (100%)

#### EXAMPLE 19

#### **Synthesis of Rapamycin-42-O-ester-(S) (-)-2-amino-3-phenyl-1-propanol (7j)**

15 The active ester 42-O-(4-Nitrophenoxy carbonyl) rapamycin (6) 250 mg (0.231 mmol) was dissolved in dry N,N-dimethylformamide (10 mL) and to it 50µL dry pyridine was added, the reaction mixture was stirred under nitrogen for five minutes at 25°C. To this stirred  
20 solution 38.5 mg (0.225 mmol) of (S) (-)-2-amino-3-phenyl-1-propanol dissolved in 1 mL of N,N-dimethylformamide was added over a period of 10 minutes with constant stirring. Stirring under nitrogen was continued for 18 h at 25°C. The progress of the reaction  
25 was monitored by TLC (ethyl acetate, R<sub>f</sub> 0.5) and mass spectrum. After 18 h of the stirring the reaction mixture was evaporated to dryness. The crude mixture was purified

on silica gel (Silica gel 60, 63-200 $\mu$ ) column chromatography, by step gradient from 10 to 50% of ethyl acetate: hexane and then pure ethyl acetate to give Rapamycin-42-O-ester-(S) (-)-2-amino-3-phenyl-1-propanol (7j) as beige colored solid 150 mg (60% yield), LC/MS showed  $M^+ Na$  1113.8 (100%) and  $M-1$  1069.8 (100%).

#### EXAMPLE 20

#### **Synthesis of Rapamycin-42-O-ester-(R) (+)-2-amino-3-phenyl-1-propanol (7k)**

10 The active ester 42-O-(4-Nitrophenoxy carbonyl) rapamycin (6) 125 mg (0.115 mmol) was dissolved in dry N, N-dimethylformamide (10 mL) and to it 50 $\mu$ L dry pyridine was added, the reaction mixture was stirred under nitrogen for five minutes at 25°C. To this stirred  
15 solution 19.2 mg (0.127 mmol) of (R) (+)-2-amino-3-phenyl-1-propanol dissolved in 1 mL of N, N-dimethylformamide was added over a period of 10 minutes with constant stirring. Stirring under nitrogen was continued for 18 h at 25°C. The progress of the reaction  
20 was monitored by TLC (ethyl acetate,  $R_f$  0.6) and mass spectrum. After 18 h of the stirring the reaction mixture was evaporated to dryness. The crude mixture was purified on silica gel (Silica gel 60, 63-200 $\mu$ ) column chromatography, by step gradient from 10 to 50% of ethyl  
25 acetate: hexane and then pure ethyl acetate to give Rapamycin-42-O-ester-(R) (+)-2-amino-3-phenyl-1-propanol (7k) as beige colored solid 65 mg (51% yield), LC/MS showed  $M-1$  1069.8 (100%).

EXAMPLE 21**Synthesis of Rapamycin-42-O-ester-(1S,2S)-(+) -2-amino-1-phenyl-1,3-propandiol (71)**

The active ester 42-O-(4-Nitrophenoxy carbonyl) rapamycin (6) 150 mg (0.139 mmol) was dissolved in dry N, N-dimethylformamide (10 mL) and to it 50µL dry pyridine was added, the reaction mixture was stirred under nitrogen for five minutes at 25°C. To this stirred solution 25.5 mg (0.139 mmol) of (1S,2S)-(+) -2-amino-1-phenyl-1,3-propandiol dissolved in 1 mL of N, N-dimethylformamide was added over a period of 10 minutes with constant stirring. Stirring under nitrogen was continued for 18 h at 25°C. The progress of the reaction was monitored by TLC (ethyl acetate,  $R_f$  0.4) and mass spectrum. After 18 h of the stirring the reaction mixture was evaporated to dryness. The crude mixture was purified on silica gel (Silica gel 60, 63-200µ) column chromatography, by step gradient from 10 to 50% of ethyl acetate: hexane and then pure ethyl acetate to give Rapamycin-42-O-ester-(1S,2S)-(+) -2-amino-1-phenyl-1,3-propandiol (71) as white colored solid 62.3 mg (41% yield), LC/MS showed M-1 1105.7 (100%).

EXAMPLE 22**Synthesis of Rapamycin-42-O-ester-2-amino-3-methyl-1-pentanol (7m)**

The active ester 42-O-(4-Nitrophenoxy carbonyl) rapamycin (6) 100 mg (0.092 mmol) was dissolved in dry N, N-dimethylformamide (6 mL) and to it 50µL dry pyridine

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was added, the reaction mixture was stirred under nitrogen for five minutes at 25°C. To this stirred solution 11.8 mg (0.101 mmol) of 2-amino-3-methyl-1-pentanol dissolved in 1 ml of N, N-dimethylformamide was added over a period of 10 minutes with constant stirring. Stirring under nitrogen was continued for 18 h at 25°C. The progress of the reaction was monitored by TLC (ethyl acetate,  $R_f$  0.6) and mass spectrum. After 18 h of the stirring the reaction mixture was evaporated to dryness. The crude mixture was purified on silica gel (Silica gel 60, 63-200 $\mu$ ) column chromatography, by step gradient from 10 to 50% of ethyl acetate: hexane and then pure ethyl acetate to give Rapamycin-42-O-ester-2-amino-3-methyl-1-pentanol (**7m**) as white colored solid 50 mg (52% yield), LC/MS showed M-1 1055.7 (100%).

### EXAMPLE 23

#### **Synthesis of Rapamycin-42-O-ester-3-amino-1, 2-propanediol (**7n**)**

The active ester 42-O-(4-Nitrophenoxy carbonyl) rapamycin (**6**) 100 mg (0.092 mmol) was dissolved in dry N, N-dimethylformamide (5 mL) and to it 50 $\mu$ L dry pyridine was added, the reaction mixture was stirred under nitrogen for five minutes at 25°C. To this stirred solution 9.29 mg (0.102 mmol) of 3-amino-1, 2-propanediol dissolved in 1 ml of N, N-dimethylformamide was added over a period of 10 minutes with constant stirring. Stirring under nitrogen was continued for 18 h at 25°C. The progress of the reaction was monitored by TLC (10% MeOH: CH<sub>2</sub>Cl<sub>2</sub>,  $R_f$  0.4) and mass spectrum. After 18 h of the



stirring the reaction mixture was evaporated to dryness. The crude mixture was purified on silica gel (Silica gel 60, 63-200 $\mu$ ) column chromatography, by step gradient from pure CH<sub>2</sub>Cl<sub>2</sub> to 10% MeOH : CH<sub>2</sub>Cl<sub>2</sub> to give Rapamycin-42-O-  
5 ester-3-amino-1,2-propanediol (**7n**) as white colored solid 30 mg (31% yield), LC/MS showed M-1 1029.6 (100%).

#### EXAMPLE 24

#### **Synthesis of Rapamycin-42-O-ester-2-amino-1, 3-propanediol (**7o**)**

10 The active ester 42-O-(4-Nitrophenoxy carbonyl) rapamycin (**6**) 100 mg (0.092 mmol) was dissolved in dry N,N-dimethylformamide (5 mL) and to it 50 $\mu$ L dry pyridine was added, the reaction mixture was stirred under nitrogen for five minutes at 25°C. To this stirred  
15 solution 9.29 mg (0.102 mmol) of 3-amino-1, 3-propanediol dissolved in 1 ml of N, N-dimethylformamide was added over a period of 10 minutes with constant stirring. Stirring under nitrogen was continued for 18 h at 25°C. The progress of the reaction was monitored by TLC (10%  
20 MeOH: CH<sub>2</sub>Cl<sub>2</sub>, R<sub>f</sub> 0.4) and mass spectrum. After 18 h of the stirring the reaction mixture was evaporated to dryness. The crude mixture was purified on silica gel (Silica gel 60, 63-200 $\mu$ ) column chromatography, by step gradient from pure CH<sub>2</sub>Cl<sub>2</sub> to 7% MeOH : CH<sub>2</sub>Cl<sub>2</sub> to give Rapamycin-42-O-  
25 ester-3-amino-1,3-propanediol (**7o**) as white colored solid 50 mg (52% yield), LC/MS showed M-1 1029.5 (100%).

EXAMPLE 25**Synthesis of Rapamycin-42-O-ester-2-amino-2-methyl-1, 3-propanediol (7p)**

The active ester 42-O-(4-Nitrophenoxy carbonyl) rapamycin (6) 100 mg (0.092 mmol) was dissolved in dry N,N-dimethylformamide (5 mL) and to it 50µL dry pyridine was added, the reaction mixture was stirred under nitrogen for five minutes at 25°C. To this stirred solution 9.7 mg (0.092 mmol) of 2-amino-2-methyl-1, 3-propanediol dissolved in 1 ml of N, N-dimethylformamide was added over a period of 10 minutes with constant stirring. Stirring under nitrogen was continued for 18 h at 25°C. The progress of the reaction was monitored by TLC (10% MeOH: CH<sub>2</sub>Cl<sub>2</sub>, R<sub>f</sub> 0.6) and mass spectrum. After 18 h of the stirring the reaction mixture was evaporated to dryness. The crude mixture was purified on silica gel (Silica gel 60, 63-200µ) column chromatography, by step gradient from pure CH<sub>2</sub>Cl<sub>2</sub> to 7% MeOH : CH<sub>2</sub>Cl<sub>2</sub> to give Rapamycin-42-O-ester-2-amino-2-methyl-1,3-propanediol (7p) as white colored solid 31 mg (33% yield), LC/MS showed M<sup>+</sup>+ Na 1067.5 (100%).

EXAMPLE 26**Synthesis of Rapamycin-42-O-ester-(2S, 3S)-2-amino-1, 3-butanediol (7q)**

25 The active ester 42-O-(4-Nitrophenoxy carbonyl)rapamycin (6) 100 mg (0.092 mmol) was dissolved in dry N,N-dimethylformamide (20 mL) and to it 50µL dry pyridine was added, the reaction mixture was

stirred under nitrogen for five minutes at 25°C. To this stirred solution 9.7 mg (0.092 mmol) of (2S, 3S)-2-amino-1, 3-butanediol dissolved in 1 ml of N, N-dimethylformamide was added over a period of 10 minutes with constant stirring. Stirring under nitrogen was continued for 18 h at 25°C. The progress of the reaction was monitored by TLC (10% MeOH: CH<sub>2</sub>Cl<sub>2</sub>, R<sub>f</sub> 0.6) and mass spectrum. After 18 h of the stirring the reaction mixture was evaporated to dryness. The crude mixture was purified on silica gel (Silica gel 60, 63-200μ) column chromatography, by step gradient from pure CH<sub>2</sub>Cl<sub>2</sub> to 7% MeOH : CH<sub>2</sub>Cl<sub>2</sub> to give Rapamycin-42-O-ester-(2S, 3S)-2-amino-1, 3-butanediol (**7q**) as beige colored solid 60 mg (62% yield), LC/MS showed M<sup>+</sup> Na 1067.7 (100%).

#### 15 EXAMPLE 27

##### **Synthesis of Rapamycin-42-O-ester-(2R, 3R)-2-amino-1, 3-butanediol (**7r**)**

The active ester 42-O-(4-Nitrophenoxy carbonyl) rapamycin (**6**) 100 mg (0.092 mmol) was dissolved in dry N, N-dimethylformamide (15 mL) and to it 50μL dry pyridine was added, the reaction mixture was stirred under nitrogen for five minutes at 25°C. To this stirred solution 9.7 mg (0.092 mmol) of (2R, 3R)-2-amino-1, 3-butanediol dissolved in 1 ml of N, N-dimethylformamide was added over a period of 10 minutes with constant stirring. Stirring under nitrogen was continued for 18 h at 25°C. The progress of the reaction was monitored by TLC (10% MeOH: CH<sub>2</sub>Cl<sub>2</sub>, R<sub>f</sub> 0.5) and mass spectrum. After 18 h of the stirring the reaction mixture was evaporated to

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dryness. The crude mixture was purified on silica gel (Silica gel 60, 63-200 $\mu$ ) column chromatography, by step gradient from pure CH<sub>2</sub>Cl<sub>2</sub> to 10% MeOH : CH<sub>2</sub>Cl<sub>2</sub> to give Rapamycin-42-O-ester-(2R, 3R)-2-amino-1, 3-butanediol (7r) as white colored solid 65 mg (67% yield), LC/MS showed M- 1 1043.6 (100%).

#### EXAMPLE 28

#### **Synthesis of Rapamycin-42-O-ester-(R)-(-)-2-amino-4-methyl pentanol (7s)**

10 The active ester 42-O-(4-Nitrophenoxy carbonyl) rapamycin (6) 100 mg (0.092 mmol) was dissolved in dry N, N-dimethylformamide (5 mL) and to it 50  $\mu$ L dry pyridine was added, the reaction mixture was stirred under nitrogen for five minutes at 25°C. To this stirred  
15 solution 10.8 mg (0.092 mmol) of (R)-(-)-2-amino-4-methyl pentanol dissolved in 1 mL of N, N-dimethylformamide was added over a period of 10 minutes with constant stirring. Stirring under nitrogen was continued for 18 h at 25°C. The progress of the reaction was monitored by TLC (10%  
20 MeOH: CH<sub>2</sub>Cl<sub>2</sub>, R<sub>f</sub> 0.5) and mass spectrum. After 18 h of the stirring the reaction mixture was evaporated to dryness. The crude mixture was purified on silica gel (Silica gel 60, 63-200 $\mu$ ) column chromatography, by step gradient from pure CH<sub>2</sub>Cl<sub>2</sub> to 5% MeOH : CH<sub>2</sub>Cl<sub>2</sub> to give Rapamycin-42-O-  
25 ester-(R)-(-)-2-amino-4-methyl pentanol (7s) as white colored solid 34 mg (35% yield), LC/MS showed M<sup>+</sup> + Na 1079.7 (100%).

EXAMPLE 29**Synthesis of Rapamycin-42-O-ester-(S)-(+) -2-amino-4-methyl pentanol (7t)**

The active ester 42-O-(4-Nitrophenoxy carbonyl) rapamycin (6) 100 mg (0.092 mmol) was dissolved in dry N, N-dimethylformamide (5 mL) and to it 50µL dry pyridine was added, the reaction mixture was stirred under nitrogen for five minutes at 25°C. To this stirred solution 10.8 mg (0.092 mmol) of (S)-(+) -2-amino-4-methyl pentanol dissolved in 1 ml of N, N-dimethylformamide was added over a period of 10 minutes with constant stirring. Stirring under nitrogen was continued for 18 h at 25°C. The progress of the reaction was monitored by TLC (10% MeOH: CH<sub>2</sub>Cl<sub>2</sub>, R<sub>f</sub> 0.5) and mass spectrum. After 18 h of the stirring the reaction mixture was evaporated to dryness. The crude mixture was purified on silica gel (Silica gel 60, 63-200µ) column chromatography, by step gradient from pure CH<sub>2</sub>Cl<sub>2</sub> to 5% MeOH : CH<sub>2</sub>Cl<sub>2</sub> to give Rapamycin-42-O-ester-(S)-(+) -2-amino-4-methyl pentanol (7t) as white colored solid 43 mg (40% yield), LC/MS showed M<sup>+</sup> + Na 1079.7 (100%).

EXAMPLE 30**Synthesis of Rapamycin-42-O-ester-Tris(hydroxymethyl) amino methane (7u)**

The active ester 42-O-(4-Nitrophenoxy carbonyl) rapamycin (6) 100 mg (0.092 mmol) was dissolved in dry N, N-dimethylformamide (5 mL) and to it 50µL dry pyridine was added, the reaction mixture was stirred under

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nitrogen for five minutes at 25°C. To this stirred solution 12.35 mg (0.102 mmol) of tris(hydroxymethyl)amino methane dissolved in 1 ml of N, N-dimethylformamide was added over a period of 10 minutes with constant stirring. Stirring under nitrogen was continued for 18 h at 25°C. The progress of the reaction was monitored by TLC (10% MeOH: CH<sub>2</sub>Cl<sub>2</sub>, R<sub>f</sub> 0.5) and mass spectrum. After 18 h of the stirring the reaction mixture was evaporated to dryness. The crude mixture was purified on silica gel (Silica gel 60, 63-200μ) column chromatography, by step gradient from pure CH<sub>2</sub>Cl<sub>2</sub> to 5% MeOH : CH<sub>2</sub>Cl<sub>2</sub> to give Rapamycin-42-O-ester-tris(hydroxymethyl)amino methane (7u) as white colored solid 25 mg (30% yield), LC/MS showed M<sup>+</sup> + Na 1083.6 (100%).

#### EXAMPLE 31

##### **Synthesis of Rapamycin-42-O-(3-carboxy propanoyl) ester-(S) (-)-2-amino-3-phenyl-1-propanol (7v)**

The active ester 42-O-(3-carboxy propanoyl) rapamycin (WO 94/24304) 75 mg (0.067 mmol) was dissolved in dry N, N-dimethylformamide (5 mL) and to it 40μL dry pyridine was added, the reaction mixture was stirred under nitrogen for five minutes at 25°C. To this stirred solution 11.1 mg (0.074 mmol) of (S) (-)-2-amino-3-phenyl-1-propanol dissolved in 1 ml of N, N-dimethylformamide was added over a period of 10 minutes with constant stirring. Stirring under nitrogen was continued for 18 h at 25°C. The progress of the reaction was monitored by TLC (5% MeOH: CH<sub>2</sub>Cl<sub>2</sub>, R<sub>f</sub> 0.6) and mass

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spectrum. After 18 h of the stirring the reaction mixture was evaporated to dryness. The crude mixture was purified on silica gel (Silica gel 60, 63-200 $\mu$ ) column chromatography, by step gradient from pure CH<sub>2</sub>Cl<sub>2</sub> to 4% MeOH : CH<sub>2</sub>Cl<sub>2</sub> to give Rapamycin-42-O-(3-carboxy propanoyl) ester-(S) (-)-2-amino-3-phenyl-1-propanol (**7v**) as white colored solid 50 mg (50% yield), LC/MS showed M<sup>+</sup> + Na 1145.4 (100%).

The embodiment(s) of the invention described above is(are) intended to be exemplary only. The scope of the invention is therefore intended to be limited solely by the scope of the appended claims.